

A Review on Potential Drug Delivery System as a Treatment of Intercellular Bacterial Infection

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Abstract

Date Received: 20/10/2020 Date Revised: 17/11/2020 Date Accepted: 23/11/2020	<p>Intracellular bacterial pathogens are hard to treat because of the inability of conventional antimicrobial agents belonging to widely used classes, like aminoglycosides and β-lactams, fluoroquinolones, or macrolides to penetrate, accumulate, or be retained in the mammalian cells. The increasing problem of antibiotic resistance complicates more the treatment of the diseases caused by these agents. In many cases, the increase in therapeutic doses and treatment duration is accompanied by the occurrence of severe side effects. Taking into account the huge financial investment associated with bringing a new antibiotic to the market and the limited lifetime of antibiotics, the design of drug delivery systems to enable the targeting of antibiotics inside the cells, to improve their activity in different intracellular niches at different pH and oxygen concentrations, and to achieve a reduced dosage and frequency of administration could represent a prudent choice. An ideal drug delivery system should possess several properties, such as antimicrobial activity, biodegradability, and biocompatibility, making it suitable for use in biomedical and pharmaceutical formulations. This approach allow reviving old antibiotics rendered useless by resistance or toxicity, rescuing the last line therapy antibiotics by increasing the therapeutic index, widening the antimicrobial spectrum of antibiotics scaffolds that failed due to membrane permeability problems, and thus reducing the gap between increasingly drug-resistant pathogens and the development of new antibiotics. Different improved drug carriers have been developed for treating intracellular pathogens, including antibiotics loaded into liposomes, microspheres, polymeric carriers, and nanoplexes. The purpose of this chapter is to present the limitations of each class of antibiotics in targeting intracellular pathogens and the main research directions for the development of drug delivery systems for the intracellular release of antibiotics.</p>
Keywords Intercellular Bacteria, Drug Delivery System, Drug Carriers, Liposomes	
<div style="display: flex; justify-content: space-between; align-items: center;"> <div data-bbox="167 1332 391 1388">  </div> <div data-bbox="486 1332 1428 1377"> <p>An official publication of Global Pharmacovigilance Society; Published under licence of Creative Commons Attribution 4.0 International COPYRIGHT © 2020 Author(s)</p> </div> </div>	

Introduction

Contaminations with intracellularly dwelling bacterial microorganisms are onerous to treat because various surprising difficulties are introduced by them. Then again, some variety of this bacterium are able to escape from the bactericidal components of cells by produce a “silent” infection inside the cells. Thus, in these circumstances, cells are bumbling to suppress the growth of intracellular microbes as well as may contribute to the proliferation of the infection to different organs and cells by acting as reservoir (Butts *et al.*, 2008). In addition, the intracellular locations of these bacterial pathogens are shielding them from host defense mechanism and from the activity of antibiotics, which may experience troubles in penetrating phagocytic cells (Abeylath *et al.*, 2008). In this manner, an enormous assortment of anti-toxins shows frustrating

outcomes against these microbes in the intracellular climate, which have been discovered to be dynamic against secluded microorganisms. Need of durable consolidated helpful regimens are emerging because of the low achievement pace of anti-infection treatment against the contaminations brought about by intracellular microorganisms. Nonetheless, albeit joined treatments are more powerful than single ones, their satisfaction turns out to be harder for patients and this can prompt a helpless patient consistence and relinquishing of the anti-infection treatment. Taken together, every one of these elements may add to the event of relapses after treatment. in reverse hand, development of a new antibiotic is associated with the massive financial investment and lifetime of the antibiotic is limited.

For eradicating the above problems an advanced drug delivery system must be developed which able to targets

antibiotics in intracellular spaces, improve their activity in various intracellular niches at various oxygen concentrations and pH, with a less frequent and considerable low dose of administration (The National Academies Press. 2010). This methodology will permit resuscitating old antibiotics delivered futile by toxicity or resistance, protecting the last line treatment antibiotics by expanding their therapeutic index, extending the antimicrobial range of antibiotics platforms that failed because of film penetrability issues, and accordingly decreasing the gap between the improvement of new antibiotics and progressively drug resistant microorganisms. In this context, the principle reason for this review is to introduce the conceivable utility of antibiotics containing drug delivery System for the treatment of intracellular diseases.

Lifestyles of intracellular bacterial pathogens

A significant specialty to be considered in the zone of infectious illness is the intracellular environment of the expert phagocytes. Strangely, these particular cells customized to demolish and digest the ingested material are a fantastic territory for bacterial microorganisms, for example, Brucella, Listeria, Salmonella or Mycobacterium. The solid developmental pressure for surviving the challenge of lethal immune system has brought about an assorted collection of microbial

techniques to make a safe replicative specialty. Facultative as well as obligate intracellular microbes present diverse virulence factors that will adjust their intracellular destiny (Alonso *et al.*, 2004). Take-up of nutrients and reusing of proteins which are utilized in the secretory pathway, is performed by endocytic pathway. Phagosomes containing ingested microbial organisms combine with early endosomes, gaining markers that present on phagosomes properties typically allotted to early endosomes, including the capacity to meld with other endocytic organelles (Desjardins, Beron *et al.*, 1995). They fuse consecutively with matured or expanding aged endosomes (lysosomes and late endosomes) securing new markers and a more acidic climate (pH 5.5). Changes in the composition of phospholipid is additionally seen during phagosome development (Desjardins *et al.*, 1995). Intracellular microorganisms have discovered approaches to redirect their standard trafficking from early endosomes towards these unfriendly phagolysosomes. Microbes would thus be able to elude from the cytoplasmic endocytic vacuole, live inside the phagolysosome, or make their own advantaged specialty (Suzuki *et al.*, 2001). Notwithstanding the methods by which this is accomplished, microorganisms can endure and repeat undetected inside the ensured environment of the host cell. Paradigmatic models require for intracellular endurance systems, is represented by few microorganisms.

Table 1 – List of some intracellular microorganisms which are able to persist inside phagocytic cells.

Name of organism	Variety of parasite	Subcellular localization	pH	References,Year
<i>Listeria monocytogens</i>	Facultative	Cytosol	~6.5	Portnoy <i>et al.</i> ,2002
<i>Shigella spp.</i>	Facultative	Cytosol	~6.5	Suzuki <i>et al.</i> ,2001
<i>Chlamydia spp.</i>	Strict	Inclusions	>6.0	Heinzen <i>et al.</i> , 1996
<i>Mycobacterium avium</i>	Facultative	Early endosomes	(5.6-6.3)	Sturgill-Koszycki <i>et al.</i> ,1996
<i>Mycobacterium bovis</i>	Facultative	Early endosomes	5.5	Hackam <i>et al.</i> , 1998
<i>Ehrlichia chaffeensis</i>	Strict	Early endosomes	Not determined	Barnewall <i>et al.</i> , 1997
<i>Coxiella burnetii</i>	Strict	Phagolysosomes	~5	Ghigo <i>et al.</i> , 2002
<i>Yersinia pseudotuberculosis</i>	Facultative	Phagolysosomes	6	Tsukano <i>et al.</i> , 1999
<i>Francisella tularensis</i>	Facultative	Phagolysosomes	6.7	Clemens <i>et al.</i> , 2004
<i>Staphylococcus aureus</i>	Facultative	Phagolysosomes	Not determined	Kubica <i>et al.</i> , 2008

Lysis of the intracellular vacuole and escape to the cytosol

Shigella and *Listeria* are instances of microbes which are able to escape from the phagocytic vacuole evading introduction to the abusive compartments of the endometrium course, and in this manner are at long last found at supplement rich host cytosol and the neutral pH. Due to presence of some phospholipases and a pore-forming hemolysin (listeriolysin O), *Listeria monocytogenes* may invade and persist into phagocytic as well as non-phagocytic cells and it attacks the cytosol (Bonazzi *et al.*, 2006). On account of *Shigella flexneri* a cytotoxin lyses the film of the phagosome and permits it to run away to the cytosol. Like *Listeria*, *Shigella* misuses the host actin polymerization apparatus and actuates the

development of actin-rich comet tails by methods for a protein called IcsA (Goldberg *et al.*, 1995).

Arrest of phagosome maturation

some pathogenic microorganisms, for example, Salmonella and Mycobacterium capture the development of the phagosome at explicit phases of the phagolysosomal route and advance into an intracellular life cycle including avoidance of phagosome-lysosome combination. As an outcome, the pH of the last confinement of the microorganisms is near to neutrality. Salmonella typhimurium momentarily secures early endosome markers yet it has created mechanisms to adjust the reallocation of lysosomal markers and endosomal and it appears to be that it might tweak the outflow of vacuolar ATPases to create a moderately less acidic phagosomal compartment (Hashim *et al.*, 2000).

Proficient phagocytes are the favored focuses of *Mycobacterium* spp. Immediately after its phagocytosis, *Mycobacterium* containing vacuoles gain a large number of early endosomal markers however not late endosomal/lysosomal ones, recommending that *Mycobacterium* capture development before the arrangement of late phagosomes (Sturgill-Koszycki et al., 1996).

Segregation from the endocytic route

Legionella, *Chlamydia* and *Brucella* phagosomes isolate from the endocytic pathway to make an appropriate environment for intracellular replication.

Legionella pneumophila is a facultative intracellular microbe that colonizes essentially alveolar macrophages. Vacuoles holding *L. pneumophila* have been demonstrated not to procure lysosomal and endosomal markers (Joshi et al., 2001) yet to enlist. Endoplasmic reticulum (ER)-determined vesicles, showing that the microscopic organisms' dwell in a compartment encompassed by ER until the monocytes lyse delivering the microorganisms (Roy et al., 2002).

The *chlamydiae* are a type of obligate intracellular microbes that fill in cytoplasmic vesicles in a wide assortment of host cells, including tissue macrophages and blood coursing monocytes. *Chlamydia* isolates from the endocytic pathway at a beginning phase of advancement and occupies membrane bound vacuoles named inclusions. No marker of the endocytic course is gathered on or inside the inclusion (Scidmore et al., 2003). These inclusions confine near the network of Golgi and wire to sphingomyelin containing exocytic vesicles, which is essentially required for the intracellular replication of *Chlamydia* (Van Ooij et al., 2002).

Brucella spp. are considered as facultative intracellular microbes which are able to contaminate and imitate inside the organs and cells or the mononuclear phagocytic framework (MPS), essentially in the spleen and the liver. At the point when they are caught by phagocytic cells, these living beings present a fantastic model for running away to a safe replicative environment undermining the ordinary phagosome development process.

Life within acidic phagosomes

Staphylococcus aureus is a pervasive bacterium. local infections which are generally created by this organism, spreads to different tissues and organs. Despite the fact that it has been viewed as an extracellular microorganism, it effectively attacks professional as well as non-professional phagocytes.

S. aureus endures in the phagolysosomal compartment of phagocytic cells and under these conditions can endure a few days prior to actuating their lysis (Kubica et al., 2002). This specific restriction of the bacterium is by all accounts the purpose behind the repetitive character of staphylococcal diseases, just as for the disappointments of

clinical medicines utilizing anti-infection agents that are dynamic in vitro.

Coxiella burnetii, an obligate intracellular microbe, set a new example of intracellular adaptation by inhabiting an acidified lysosomal-like compartment (Bonazzi et al., 2006). It has been shown that after its entrance into human macrophages *Coxiella* acquires late endosomal-early lysosomal markers and resides in acidic vacuoles with pH 5, remaining metabolically active.

Efficiency of different classes of against intracellular pathogens

Acquiring a potential concentration of a given medication inside the target cell isn't sufficient to accomplish the destruction of intracellular microorganism. Inside cells, intracellular microorganisms are not just shielded from the extracellular climate (supplement, antibodies, and even a few anti-microbials),

yet additionally these new physicochemical intracellular conditions instigate key (metabolic and auxiliary) changes, making them impervious to anti-microbials that are dynamic in vitro. The antibiotics which can satisfy a progression of criteria, including the capacity to invade and persist by the cell, the ability to arrive at the intracellular objective, and the presentation of action in that exceptional climate where the microbes live, can deliver a fruitful treatment.

Beta-lactams and aminoglycosides- These antibiotics can't accomplish high intracellular concentration due to their inefficiency of infiltrating cells, consequently presents poor intracellular action. However, antibiotics of these class shows potential activity against facultative intracellular pathogens, including *Mycobacterium tuberculosis*.

Beta-lactam antibiotics hinders peptidoglycan synthesis for exerting their antimicrobial action. In spite of being weakly acidic, the greater variety of the beta-lactams have the capability of invading biologic films, don't accumulate inside the cells, most likely due both to the more acidic nature of the cell cytosol contrasted with the extracellular milieu and dynamic anti-infection efflux siphons (Van Bambeke et al., 2003). Nonetheless, it has been demonstrated that the low concentration of beta lactams in cells can be remunerated by their great action at acidic pH, giving these medications remedial potential regardless of their clearly troublesome cell pharmacokinetics (Barcia-Macay et al., 2006).

Aminoglycosides show a restricted intracellular action contrasted with their high bactericidal movement in an extracellular medium. These medications diffuse gradually and ineffectively through cell films because of their high hydrophilicity, yet they might be fused into macrophages utilizing a liquid stage pinocytosis measure when applied at high fixations and long incubation times (Carrin et al., 2002). As opposed to the beta-lactams, aminoglycosides are weakly basic, so in the wake of arriving at the inside of

the cell they are kept to the lysosomes, where the acidic pH may smother their action. This absence of movement has been identified with the protonation of the atom at acidic pH. Since aminoglycosides enter the microorganisms by dynamic vehicle, factors influencing this vehicle would lessen their antibacterial action (Mingeot-Leclercq *et al.*, 2002). Another significant issue that restricts the utilization of aminoglycosides is their notable toxicity. The fundamental adverse impact of aminoglycosides is nephrotoxicity, because of renal cortical collection, followed by ototoxicity and, less significantly, neuromuscular blockade (Mingeot-Leclercq *et al.*, 1999).

Macrolides and Quinolones – Rather than Beta-lactams and aminoglycosides are, this class of antibiotics are known to be dynamic against obligate intracellular and facultative intracellular organisms, due to their high cellular aggregation. Be that as it may, their intracellular action and accumulation doesn't have an immediate relationship consistently. Inside the cells macrolides circulate basically to the cytosol and lysosomes where they are caught by protonation. Notwithstanding, the final result relies upon the macrolide utilized. A few examinations have indicated that azithromycin, for example, might be basically bacteriostatic against *S. aureus* (Seral *et al.*, 2003) and *Listeria monocytogenes* (Van Ooij *et al.*, 2000), presumably in light of the fact that their acidic pH climate diminishes macrolides action (Domingo *et al.*, 1995).

Quinolones, another class of anti-toxins that accomplish high intracellular fixations in both contaminated and non-contaminated cells, can diffuse to different subcellular compartments without accumulation with a particular cell structure (Carlier *et al.*, 1990). Most of the quinolones are powerful against intracellular microorganisms as *Listeria monocytogenes* (Van Ooij *et al.*, 2002) and *Legionella* (Baltch *et al.*, 2005) however some of them show diminished action at lower pH, which would clarify the disparity between their extracellular and intracellular exercises while treating microorganisms that are restricted inside phagolysosomes (Nguyen *et al.*, 2006).

Gentamicin - At long last, it has been set up that antimicrobial medications can impact the cooperation among microorganisms and phagocytes (Cuffini *et al.*, 2004). Antibiotics as a rule can change different elements of the antimicrobial action of the phagocytic cells of the host. For instance, gentamicin upgrades the intracellular obliteration of microorganisms by the macrophage (Drevets *et al.*, 1994), while the different aminoglycosides hinder this capacity (Labro *et al.*, 2000). In actuality, chemotaxis, oxidative burst or cytokine creation are not influenced by aminoglycosides. At last, there are contemplations that demonstrate that this sort of antibiotics doesn't impact the endocytic apparatus, while different works recommend inhibitory impacts (Van den Broek *et al.*, 1989).

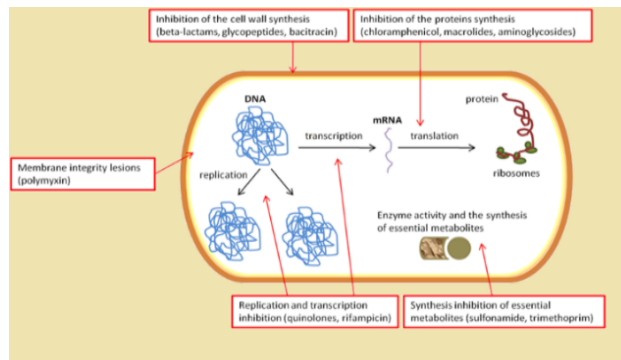


Figure 1: Cellular targets and mechanisms of action of antibiotics inside the bacterial cell.

Encapsulation of antibiotic in drug delivery systems

Improvement of new anti-microbial is certainly not a perpetual solution for treatment of intracellular diseases. The fundamental challenge for intracellular chemotherapy is to plan and build up a transporter framework for antibiotics that could be effectively endocytosed by phagocytic cells and, subsequent to attacking the cell film, ready to deliver satisfactory medication discharge. In this sense, lipidic and polymeric DDS are appropriate as vehicles for the conveyance of antimicrobial agent since they ordinarily give a sustained drug release impact and increment the general medication efficacy by limiting the toxicity related with the encapsulated drugs. Also, DDS shield the consolidated medication from untimely enzymatic and immunological assaults and, at times, they act synergistically with cell bactericidal components (Prior *et al.*, 2002).

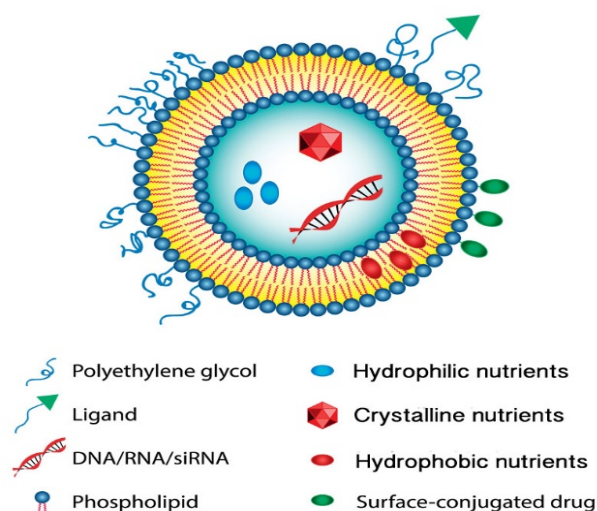


Figure 2: Schematic diagram of nanoliposome

Liposomes

Since their revelation in the mid-1960s, liposomes have been one of the most broadly examined drug transporters. They are circular vesicles of one or a few concentric lipid bilayers, typically made out of cholesterol and

phospholipids, enclosing fluid compartments (Gregoriadis *et al.*, 1976). Also, because of their comparability to organic layers they present low immunogenicity and toxicity. Fundamentally, liposomes are gaining interest due to their basic adaptability and the chance of entangling medications of diverse solvency properties and size. Contingent upon the technique utilized for their planning, liposomes of various sizes and number of lamellae can be acquired (Cordeiro *et al.*, 2000). In view of these boundaries liposomes can be grouped into large unilamellar vesicles (LUV), small unilamellar vesicles (SUV), and small plurilamellar vesicles (SPLV) (Dijkstra *et al.*, 1985). Besides, liposomes can encapsulate both hydrophilic and hydrophobic drugs. Water-solvent mixes are situated in the fluid spaces while lipid-dissolvable ones are bound or joined into the lipid film.

Liposomes poses some physicochemical properties which can be altered by changing (Deol *et al.*, 1997):

- The kinds of lipids.
- The size of the liposome.
- The arrangement and proportions of lipids in the formulation of liposome.
- Temperature affectability.
- The charge of the liposomal surface: negative, positive, or nonpartisan pH affectability.
- The fluidity of the liposomal film: inflexible and liquid liposomes.

The advantages of liposomal drug delivery

Liposomes have various advantages as antibiotic carriers which are as given below.

Improved pharmacokinetics and, biodistribution, decreased toxicity

There are a lot of proof of the advantages of liposomes as antibiotic conveyance frameworks. The benefit of liposomal transporters is the chance of a steady and continued arrival of anti-infection agents during drug dissemination in the body. This permit keeping up the legitimate medication focus for a moderately long haul. In correlation, organization of the free anti-toxin displays a speedy and short impact and requires a few dosages for each day (Gregoriadis *et al.*, 1993). Drug embodiment in liposomal vesicles improves the pharmacokinetics and furthermore ensures anti-toxins against the hydrolytic action of compounds and substance what's more, immunological deactivation. Customary liposomes applied by intravenous organization are perceived as unfamiliar antigens by the immunological framework and are opsonized. This actuates vague guard components and the liposomes are taken up by the mononuclear phagocyte framework (MPS), which prompts lower blood flow time and quick blood clearance. This property (phagocytosis of liposomes) is attractive for intracellular microbe annihilation, however ominous for different sorts of

infection (Gregoriadis *et al.*, 1993) The MPS take-up rate relies upon a few liposomal properties, for example, charge, size, and smoothness. The blood leeway of little vesicles (~100nm) ascends to a few hours, in examination with a few minutes for MLV details. Unbending and uncharged vesicles course more than liquid and charged ones. It was exhibited that expanded fine porousness at the contamination site caused a high nearby centralization of liposomes. Moreover, the presence of bacterial antigens initiated a fiery reaction, which prompted a further expansion in liposome extravagation. These systems fundamentally improved the antibacterial movement of the typified anti-infection agents and permitted the annihilation of microbe from the contaminated tissue (Hamidi 2006; Medina *et al.*, 1996)

Target selectivity: Intensive research on drug carriers demonstrated the possibility to target liposomes to particular organs, tissues, and even microorganisms (Gilbert *et al.*, 1990). Target selectivity of liposomal drug formulations may be achieved by:

- Addition of specific immunoglobulins; addition of proteins.
- Addition of specific oligosaccharide chains.
- Construction of pH-sensitive vesicles.
- Construction of thermo-sensitive vesicles, the composition of the vesicle surface conditions, the type of specific and nonspecific interaction with the target (Kohane *et al.*, 2006).

In the case of nonspecific action, the charge of the membrane plays the main role. Eukaryotic and bacterial cells possess negatively charged surfaces, which is why positively charged liposomal vesicles exhibited the strongest vesicle-cell interactions (Lasic *et al.*, 1998). Specifically, targeted liposomes are equipped with proteins, antibodies, or immunoglobulin fragments which have affinity to specific receptors located on the target surface (infected cells or pathogens). Specifically, coated vesicles could be directed toward particular infected tissue or to strictly defined pathogens. Liposomes as drug carriers are very promising in preventing biofilm formation and treatment. The cationic liposomes were more efficient than anionic in adhering to skin bacteria (Krieger *et al.*, 1999). The researchers proposed that lectin- carbohydrate interactions are the principle mechanism for drug delivery to plaque-forming bacteria.

The interactions between vesicles and epitomes expressed on the bacterial cell surface, such as glycocalyx, were studied and it was shown that polysaccharide-coated vesicles were an efficient system of metronidazole delivery to periodontal pocket biofilm and inhibition of pathogenic bacteria. The addition of zinc particles significantly increased the inhibitive effect on microbial growth and a synergic effect between the applied antimicrobials and zinc was noted. As mentioned previously, cationic formulations of liposomes exhibited

significant adhere to the skin-associated bacteria (Martel *et al.*, 2009). These are presented in the Table 2.

Table 2: Some of Liposomal antibiotics used for intracellular bacteria eradication (Jung *et al.*, 2009)

Anti-microbial Agent	Bacteria
Ampicillin	Listeria monocytogenes, Brucella abortus.
Ciprofloxacin	Salmonella enterica, Francisella tularensis.
Gentamicin	Salmonella enterica, Listeria monocytogenes.

Enhanced activity against intracellular pathogens and overcoming bacterial drug resistance

The utilization of liposomes as a medication conveyance framework was fruitful in killing intracellular microorganisms. Liposomes were applied to different kinds of diseases. In the treatment of sicknesses brought about by intracellular microbes, unbending ordinary liposomal vesicles and PEG-covered one's improved medication maintenance in the best possible tissues, given supported delivery, diminished harmfulness, and upgraded the fixation at the site of contamination. A few tests focused in on tuberculosis, a serious and hard to treat contamination. It was demonstrated that the utilization of liposomal types of rifampin, isoniazid, and clarithromycin was very effective (Hu *et al.*, 2010; Huynh *et al.*, 2009). There is likewise the chance of controlled medication discharge by utilizing pH-sensitive liposomes. Their structure is stable in the blood dissemination, however in a modified pH climate (in phagolysosomes), a flimsy layer permits spillage of medication content. The pH-touchy PEG-covered vesicles made out of lipids were applied to intracellular microorganisms, for example, *L. monocytogenes* and *Salmonella sp.* (Joralemon *et al.*, 2010).

Polymeric micro-and nanoparticles

Microparticles and nanoparticles were created as alternative frameworks to liposomes, to tackle their steadiness issues during capacity and after organization in natural liquids. This improved dependability and the chance of acquiring a modulable controlled arrival of the embodied medication are the primary favorable circumstances of polymeric particles over liposomal transporters. Because of advances in methods for microencapsulation and the improvement of new polymers, such vectors are as of now the subject of a broad examination on the exemplification of numerous dynamic fixings, for example, hormones, anticancer antigens and anti-infection agents (Mundargi *et al.*, 2008). Over the most recent couple of many years a few strategies were created to plan polymeric miniature and nanoparticles (Jain *et al.*, 2000). Some of the basic techniques utilized for polymeric molecule plan are emulsification-dissolvable evacuation (or dissolvable

vanishing), stage detachment (or coacervation), interfacial polymerization and shower drying. The choice of a specific strategy depends generally on the physicochemical qualities of the medication of interest. When all is said in done, the polymers utilized for drug exemplification purposes can be comprehensively partitioned into two gatherings, contingent upon their tendency: normal and manufactured polymers. Normal polymers (for example human or cow-like serum egg whites, gelatin, collagen, alginate, chitosan, hyaluronan, starch) are polymers gotten from regular sources. Their absence of virtue and homogeneity and the danger of sickness transmission have prompted a decline in their utilization. Then again, manufactured biodegradable and biocompatible polymers (for example poly(α -hydroxyacids), polyanhydrides like poly (sebacic corrosive) and poly (fatty corrosive dimer-sebacic corrosive), are widely utilized for the embodiment of numerous medications. Among these manufactured polymers, the aliphatic polyesters poly (lactic corrosive) (PLA), and copolymer poly (lactic co-glycolic corrosive) (PLGA) are the most broadly researched class of polymers as to toxicological and clinical information (Tewers *et al.*, 2006). PLA and PLGA are non-harmful, biocompatible furthermore, biodegradable polymers endorsed by the Food and Drug Administration for human utilization (Wakiyama *et al.*, 1982; Manson *et al.*, 1976).

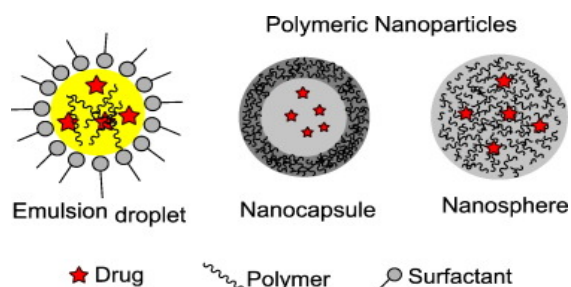


Figure 3: Various polymeric nanoparticles

Advantages of using nanomedicine

Nanomedicines offers the following advantages

Biocompatibility and nano toxicity: Biocompatibility are a significant highlight of any medication conveyance framework, and the objective is to limit vague cytotoxic impacts to solid tissues while expanding drug impacts at the objective tissue or against intrusive microbes. PLGA shows great biocompatibility, biodegradability, reasonable debasement energy, and mechanical properties and is anything but difficult to measure. For this reason, PLGA is an appealing contender for nanoparticle-based medication conveyance frameworks, and there is a huge group of progressing research in this area (Schiffelers *et al.*, 2007). Other polymers, for example, PLA, PMA, PEG, chitosan, gelatin, what's more, alginate additionally show guarantee as medication conveyance vehicles due to their biocompatible properties. Stake might be utilized as a "covering" to forestall the fast expulsion of nanoparticles from the circulation system by the mononuclear

phagocytic framework, which may increment nanoparticle dissemination time and hypothetically improve the helpful limit of the nanoparticle (Seleem *et al.*, 2009).

Cellular penetration and intracellular delivery: One fundamental challenge in treating intracellular microbes is to get enough medications to arrive at the microbe inside an intracellular compartment. Nanoparticles may live inside acidic endolysosome compartments following endocytosis, and untimely medication discharge inside this acidic compartment may cause drug debasement and render insufficient treatment. It is hence significant for the nanoparticle to get away this compartment and access the cytosol where either the medication payload might be legitimately delivered or the nanoparticle proceeds to further focus on a particular organelle. For example, PLGA nanoparticles conveying doxorubicin are purportedly fit for getting away the endolysosomal compartment by an inversion of their surface charge. This permits the particles to connect with the layer and departure into the cytosol where the doxorubicin is released (Sessa *et al.*, 1968). There are various sources that report time-and focus subordinate take-up of nanoparticles by an assortment of cell types, for example, smooth muscle cells, endothelial cells, macrophages, and tumor cells. The take-up of PLGA nanoparticles containing cow-like serum egg whites as a model medication was discovered to be focus subordinate in human vascular endothelial cells, moving toward first-request energy. An *in vitro* take-up and cell dealing study utilizing mesoporous mixture silica nanoparticles exhibited that the particles were disguised by receptor-intervene endocytosis, were confined in the endocytic compartment, and afterward delivered their load inside the cytosol (Sihorkar *et al.*, 2001).

Site-explicit and tunable medication discharge

In request to viably annihilate intracellular microorganisms, medicates that are expected to slaughter the microorganisms straightforwardly should arrive at the intracellular areas of tainted have cells. The areas may incorporate phagosomes, cytosol, vacuoles, furthermore, core and may collaborate with the endoplasmic reticulum or Golgi body. With legitimate designing and plan, nanoparticles can be custom fitted to convey their medication payloads into the contaminated cells and afterward discharge the medication inside explicit intracellular compartments. One approach to achieve site-explicit medication discharge is to utilize pH-responsive polymers (Swenson *et al.*, 1988). In one of the examinations, short peptides were formed to pH-responsive polymers planned explicitly to upset the endosomal film at pH 5.5 and thusly discharge the peptide into the cytosol. The polymers had no film problematic movement at pH 7.4 because of a "covering" PEG gathering, which is later severed to uncover the layer disturbance area at pH 5.5. Unconjugated peptide was found fundamentally in the lysosome, demonstrating that the peptide itself couldn't escape into the cytosol. This

polymer innovation may take into consideration neighborhood drug conveyance to the cytosol, despite the fact that it is more basic to arrive at the particular intracellular area of the microbe, for example, a vacuole or the nucleus (Torchilin *et al.*, 2005). It has been illustrated that nanoparticles can be explicitly focused to mitochondria or core and might be fit for entering vacuoles where microbes for example, Salmonella may dwell during a contamination cycle. Despite the fact that intracellular microbes don't commonly live inside mitochondria, pathogens, for example, *Listeria monocytogenes* can discharge poisons that meddle with typical mitochondrial work. So, the capacity to target mitochondria may give a way to treat these kinds of contamination furthermore, constrict the impacts of emitted toxins (Ulrich *et al.*, 2002).

Conclusions

Up to now, no anti-toxin treatment has been accounted for to destroy most intracellular microbes such as *Mycobacterium* and *Brucella*. Besides, a prolonged exposure to joined anti-microbials are needed to decrease the sickness backslides down to 5-15 %. In this sense, DDS have a significant part in the administration of intracellular diseases. As expressed above, detailing of antimicrobials in DDS can decrease the symptoms of these medications and increment tolerant consistence, in this way getting a good deal on wellbeing care conveyance. In this setting, the likely utilization of DDS stacked with anti-microbials might be one of the most important restoratives propels in the treatment of intracellular bacterial illnesses as of late. Liposomes have indicated great potential in improving the adequacy and decency of anti-infection agents of current use, be that as it may, issues concerning their security during capacity and organization require thorough consideration. Then again, polymeric particles, principally nanoparticles, have arisen more as of late as appealing transporters for the conveyance of medications to contaminated cells. Manufactured biodegradable and biocompatible polymers have been demonstrated to be viable for embodying an incredible assortment of anti-microbials. In addition, these polymeric particles firmly improve phagocytosis and are appropriate for intracellular conveyance of antibacterial specialists. In this manner, almost certainly, before long affirmation of the wellbeing properties of these anti-microbial stacked DDS pilot concentrates in people could be started.

References

- Abeylath SC, Turos E. Drug delivery approaches to overcome bacterial resistance to beta-lactam antibiotics. *Expert Opin Drug Delivery*. 2008;5(9):5931-5949.
- Alonso A and Garcia-del Portillo F. Hijacking of eukaryotic functions by intracellular bacterial pathogens. *Int Microbiol*, 2004;7(3), 181-191.

- Antibiotic Resistance: Implications for Global Health and Novel Intervention Strategies: Workshop Summary. Washington, DC: *The National Academies Press*. 2010, 496.
- Baltch AL, Bopp LH, Smith RP, Michelsen PB and Ritz WJ. Antibacterial activities of gemifloxacin, levofloxacin, gatifloxacin, moxifloxacin and erythromycin against intracellular *Legionella pneumophila* and *Legionella micdadei* in human monocytes. *J Antimicrob Chemother*, 2005; 56(1):104-109.
- Barcia-Macay M, Seral C, Mingeot-Leclercq MP, Tulkens PM and Van Bambeke F. Pharmacodynamic Evaluation of the Intracellular Activities of Antibiotics against *Staphylococcus aureus* in a Model of THP-1 Macrophages. *Antimicrob. Agents Chemother*. 2006, 50(3):841-851.
- Barnewall RE, Rikihisa Y and Lee EH. Ehrlichia chaffeensis inclusions are early endosomes which selectively accumulate transferrin receptor. *Infect Immun*, 1997; 65(4):1455-1461.
- Beron W, Alvarez-Dominguez C, Mayorga L and Stahl PD. Membrane trafficking along the phagocytic pathway. *Trends in Cell Biology*, 1995; 5(3):100-104.
- Bonazzi M and Cossart P. Bacterial entry into cells: a role for the endocytic machinery. *FEBS Lett*, 2006; 580(12): 2962-2967.
- Butts JD. Intracellular concentrations of antibacterial agents and related clinical implications. *Clin Pharmacokinet*, 1994;27(1):63-84.
- Carlier MB, Scorneaux B, Zenebergh A, Desnottes JF and Tulkens PM. Cellular uptake, localization and activity of fluoroquinolones in uninfected and infected macrophages. *J Antimicrob Chemother*,1990, 26:27-39.
- Carryn S, Van Bambeke F, Mingeot-Leclercq MP and Tulkens PM. Comparative intracellular (THP-1 macrophage) and extracellular activities of beta-lactams, azithromycin, gentamicin, and fluoroquinolones against *Listeria monocytogenes* at clinically relevant concentrations. *Antimicrob Agents Chemother*, 2002; 46(7): 2095-2103.
- Clemens DL, Lee BY and Horwitz MA. Virulent and avirulent strains of *Francisella tularensis* prevent acidification and maturation of their phagosomes and escape into the cytoplasm in human macrophages. *Infect Immun*, 2004;72(6): 3204-3217.
- Cordeiro C, Wiseman DJ, Lutwyche P, et al. Antibacterial efficacy of gentamicin encapsulated in pH-sensitive liposomes against an *in vivo* *Salmonella enterica* serovar Typhimurium intracellular infection model. *Antimicrob Agents Chemother*, 2000;44(3):533-539.
- Cuffini AM, Tullio V, Mandras N, Roana J, Banche G and Carlone NA. The leading role of antimicrobial agents in modulating the binomial host-microorganism. *Curr. Med. Chem.-Anti-Infective Agents*, 2004;3:1-13.
- Deol P, Khuller GK. Lung specific stealth liposomes: stability, biodistribution and toxicity of liposomal antitubercular drugs in mice. *Biochim Biophys Acta*. 1997;1334(2-3):161-172.
- Desjardins M, Huber LA, Parton RG and Griffiths G. Biogenesis of phagolysosomes proceeds through a sequential series of interactions with the endocytic apparatus. *J. Cell Biol*. 1994; 124(5): 677-688.
- Desjardins M. Biogenesis of phagolysosomes: the kiss and run hypothesis. *Trends in Cell Biology*, 1995; 5(5): 183-186.
- Dijkstra J, van Galen WJM, Regts J, et al. Uptake and processing of liposomal phospholipids by Kupffer cells in vitro. *Eur J Biochem*. 1985;148(2):391-397.
- Domingo S, Gastearena I, Diaz R and Gamazo C. Significance of environmental conditions (pH and serum) on the *in vitro* potency of azithromycin against *Brucella melitensis*. *J Chemother*,1995; 7 (4): 29-31.
- Drevets DA, Canono BP, Leenen PJ and Campbell PA. Gentamicin kills intracellular *Listeria monocytogenes*. *Infect Immun*, 1994; 62(6):2222-2228.
- Ghigo E, Capo C, Tung CH, Raoult D, Gorvel JP and Mege JL. *Coxiella burnetii* survival in THP-1 monocytes involves the impairment of phagosome maturation: IFN-gamma mediates its restoration and bacterial killing. *J Immunol*, 2002; 169(8):4488-4495.
- Gregoriadis G. Engineering liposomes for drug delivery: progress and problems. *Trends Biotechnol*. 1995;13(12):527-537.
- Gregoriadis G. The carrier potential of liposomes in biology and medicine. *N Engl J Med*. 1976;295(14):765- 770.
- Hackam DJ, Rotstein OD, Zhang W, Gruenheid S, Gros P and Grinstein S. Host resistance to intracellular infection: mutation of natural resistance-associated macrophage protein 1 (Nramp1) impairs phagosomal acidification. *J Exp Med*, 1998; 188(2):351-364.
- Hamidi M, Azadi A, Rafiei P. Pharmacokinetic consequences of pegylation. *Drug Deliv*. 2006;13(6):399-409.
- Hashim S, Mukherjee K, Rajee M, Basu SK and Mukhopadhyay A. Live *Salmonella* Modulate Expression of Rab Proteins to Persist in a Specialized Compartment and Escape Transport to Lysosomes. *J. Biol. Chem.*, 2000; 275(21):16281-16288.
- Heinzen RA, Scidmore MA, Rockey DD and Hackstadt T. Differential interaction with endocytic and exocytic

- pathways distinguish parasitophorous vacuoles of *Coxiella burnetii* and *Chlamydia trachomatis*. *Infect Immun*, 1996; 64(3): 796-809.
- Hu CM, Kaushal S, Cao HST, et al. Half-antibody functionalized lipid-polymer hybrid nanoparticles for targeted drug delivery to carcinoembryonic antigen presenting pancreatic cancer cells. *Mol Pharm*. 2010;7(3):914-920.
- Huynh NT, Passirani C, Saulnier P, et al. Lipid nanocapsules: a new platform for nanomedicines. *Int J Pharm*. 2009;379(2):201-209.
- Jain RA. The manufacturing techniques of various drug loaded biodegradable poly(lactide-co-glycolide) (PLGA) devices. *Biomaterials*, 2000; 21(23): 2475-2490.
- Joralemon MJ, McRae S, Emrick T. PEGylated polymers for medicine: from conjugation to self-assembled systems. *Chem Comm*. 2010;46(9):1377-1393.
- Gilbert P, Collier PJ, Brown MRW. Influence of growth rate on susceptibility to antimicrobial agents: biofilms, cell cycle, dormancy, and stringent response. *Antimicrob Agents Chemother*. 1990;34(10):1865-1868.
- Goldberg MB and Theriot JA. *Shigella flexneri* surface protein IcsA is sufficient to direct actin-based motility. *Proc Natl Acad Sci USA*, 1995; 92(14) :6572-6576.
- Gregoriadis G. Liposome preparation and related techniques. In: Gregoriadis G, editor. *Liposome Technology*. Boca Raton: CRC Press; 1993. 1-63p.
- Gregoriadis G. The carrier potential of liposomes in biology and medicine (first of two parts). *N Engl J Med*, 1976; 295(13): 704- 710.
- Joshi AD, Sturgill-Koszycki S and Swanson MS. Evidence that Dot-dependent and independent factors isolate the *Legionella pneumophila* phagosome from the endocytic network in mouse macrophages. *Cell Microbiol*, 2001;3(2): 99-114.
- Jung SK, Lim DH, Jung SH, et al. Amphotericin B-entrapping lipid nanoparticles and their in vitro and in vivo characteristics. *Eur J Pharm Sci*. 2009;37(3-4):313-320.
- Kohane DS, Tse JY, Yeo Y, et al. Biodegradable polymeric microspheres and nanospheres for drug delivery in the peritoneum. *J Biomed Mater Res*. 2006;77(2):351-361.
- Krieger J, Childs S, Klimberg I. UTI treatment using liposomal amikacin in Berlin. *Clinical Microbiology*. 1999;5(Suppl 3):136-144.
- Kubica M, Guzik K, Koziel J, Zarebski M, Richter W, Gajkowska B, A. Golda A, A. Maciag-Gudowska A, K. Brix K, L. Shaw L, T. Foster T and J. Potempa J. A potential new pathway for *Staphylococcus aureus* dissemination: the silent survival of *S. aureus* phagocytosed by human monocyte-derived macrophages. *PLoS One*, 2008; 3(1), e1409.
- Labro MT. Interference of antibacterial agents with phagocyte functions: immunomodulation or "immuno-fairy tales"? *Clin Microbiol Rev*, 2000; 13(4): 615-650.
- Lasic DD. Novel applications of liposomes. *Trends Biotechnol*, 1998;16(7):307-321.
- Manson N, Thies C and Cicero TJ. *In vivo* and *in vitro* evaluation of a microencapsulated narcotic antagonist. *J. Pharm. Sci.*, 1976; 65: 51-59.
- Martel S. Disadvantage of nanomedicine. *Int J Nanomedicine*, 2009; 50:1- 5.
- Medina C, Rahme K, Arcy DM, et al. Poloxamer mixed micelles for delivery of gambogic. *Int J Nanomedicine*, 1996; 10:407-409
- Mingeot-Leclercq MP and Tulkens PM. Aminoglycosides: Nephrotoxicity. *Antimicrob. Agents Chemother*, 1999; 43(5): 1003- 1012.
- Mingeot-Leclercq MP, Glupczynski Y and Tulkens PM. Aminoglycosides: Activity and Resistance. *Antimicrob. Agents Chemother.*, 1999; 43(4) :727-737.
- Mundargi RC, Babu VR, Rangaswamy V, Patel P and Aminabhavi TM. Nano/micro technologies for delivering macromolecular therapeutics using poly(D, L-lactide-co-glycolide) and its derivatives. *J Control Release*, 2008; 125(3): 193-209.
- Nguyen HA, Grellet J, Paillard D, Dubois V, Quentin C and Saux MC. Factors influencing the intracellular activity of fluoroquinolones: a study using levofloxacin in a *Staphylococcus aureus* THP-1 monocyte model. *J Antimicrob Chemother*, 2006; 57(5): 883-890.
- Portnoy DA, Auerbuch V and Glomski IJ. The cell biology of *Listeria monocytogenes* infection: the intersection of bacterial pathogenesis and cell-mediated immunity. *J Cell Biol*, 2002; 158(3) :409-414.
- Prior S, Gander B, Blarer N, Merkle HP, Subira ML, Irache JM and Gamazo C. *In vitro* phagocytosis and monocytemacrophage activation with poly(lactide) and poly(lactide-co-glycolide) microspheres. *Eur J Pharm Sci*, 2002; 15(2):197-207.
- Roy CR and Tilney LG. The road less traveled: transport of *Legionella* to the endoplasmic reticulum. *J Cell Biol*, 2002; 158(3):415-419.
- Schiffelers RM, Storm G, Bakker-Woudenberg IA. Host factors influencing the preferential localization of sterically stabilized liposomes in *Klebsiella*

- pneumoniae-infected rat lung tissue. *Pharm Res.*, 2001;18(6):780–787.
- Scidmore MA, Fischer ER and Hackstadt T. Restricted fusion of Chlamydia trachomatis vesicles with endocytic compartments during the initial stages of infection. *Infect Immun.*, 2003;71(2):973-984.
- Seleem MN, Munusamy P, Ranjan A, et al. Silica-antibiotic hybrid nanoparticles for targeting intracellular pathogens. *Antimicrob Agents Chemother.* 2009; 53(10):4270-4274.
- Seral C, Van Bambeke F and Tulkens PM. Quantitative analysis of gentamicin, azithromycin, telithromycin, ciprofloxacin, moxifloxacin, and oritavancin (LY333328) activities against intracellular Staphylococcus aureus in mouse J774 macrophages. *Antimicrob Agents Chemother.* 2003; 47(7):2283-2292.
- Sessa G, Weissmann G. Phospholipid spherules (liposomes) as a model for biological membranes. *J Lipid Res.* 1968;9(3):310-318.
- Sihorkar V, Vyas SP. Biofilm consortia on biomedical and biological surfaces: delivery and targeting strategies. *Pharm Res.* 2001;18(9):1247-1254.
- Sturgill-Koszycki S, Schaible UE and Russell DG. Mycobacterium-containing phagosomes are accessible to early endosomes and reflect a transitional state in normal phagosome biogenesis. *EMBO J*, 1996; 15(24): 6960-6968.
- Suzuki T and Sasakawa C. Molecular basis of the intracellular spreading of Shigella. *Infect Immun.* 2001; 69(10) :5959-5966.
- Swenson CE, Popescu MC, Ginsberg RS. Preparation and use of liposomes in the treatment of microbial infections. *Crit Rev Microbiol.*, 1988;15(Suppl 1):S1-S31.
- Tewers F, Boury F and Benoit JP. Biodegradable Microspheres: Advances in Production Technology. In *Microencapsulation: Methods and Industrial Applications*. Ed S. Benita. Marcel Dekker, New York (2006)
- Torchilin VP. Recent advances with liposomes as pharmaceutical carriers. *Nat Rev Drug Discov.* 2005;4(2):145-160.
- Tsukano H, Kura F, Inoue S, Sato S, Izumiya H, Yasuda T and Watanabe H. Yersinia pseudotuberculosis blocks the phagosomal acidification of B10.A mouse macrophages through the inhibition of vacuolar H (+)-ATPase activity. *Microb Pathog*, 1999; 27(4): 253-263.
- Ulrich AS. Biophysical aspects of using liposomes as delivery vehicles. *Biosci Rep.* 2002;22(2):129-150.
- Van Bambeke F, Michot JM and Tulkens PM. Antibiotic efflux pumps in eukaryotic cells: occurrence and impact on antibiotic cellular pharmacokinetics, pharmacodynamics and toxicodynamics. *J Antimicrob Chemother.* 2003; 51(5):1067-1077.
- Van den Broek PJ. Antimicrobial drugs, microorganisms, and phagocytes. *Rev Infect Dis.* 1989; 11(2):213-245.
- Van Ooij C, Kalman L, van I, Nishijima M, Hanada K, Mostov K and Engel JN. Host cell-derived sphingolipids are required for the intracellular growth of Chlamydia trachomatis. *Cell Microbiol.* 2000; 2(6):627-637.
- Wakiyama N, Juni K and Nakano M. Preparation and evaluation in vitro and in vivo of polylactic acid microspheres containing dibucaine. *Chem Pharm Bull (Tokyo)*, 1982; 30(10):3719-3727.